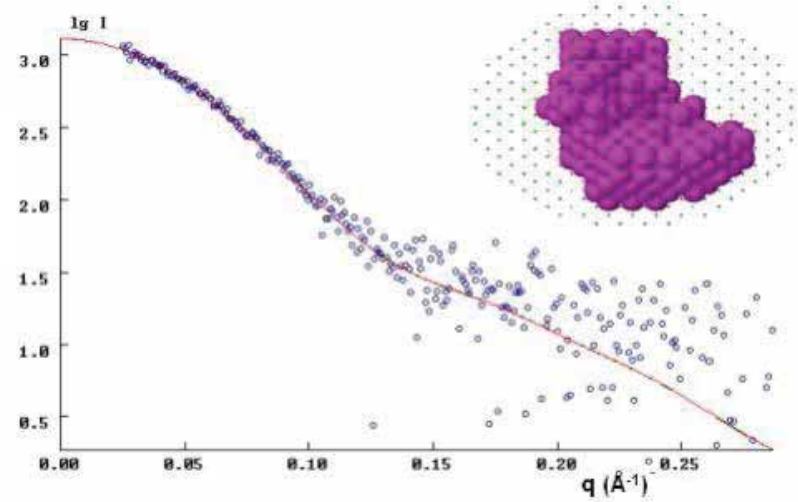
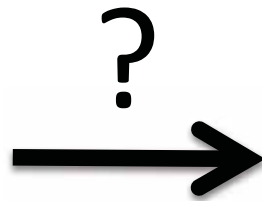
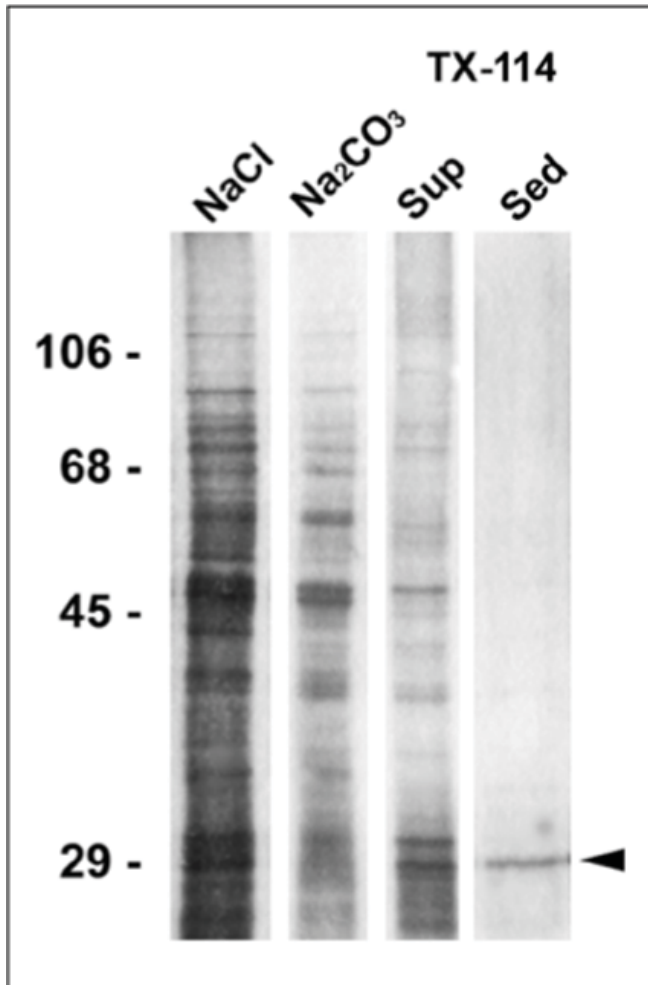


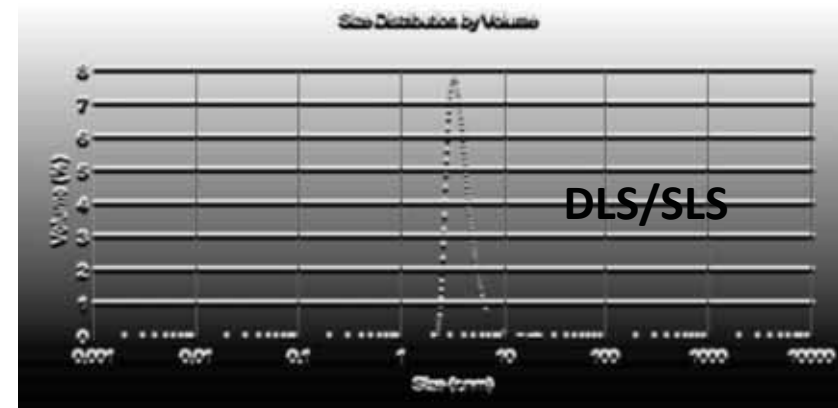
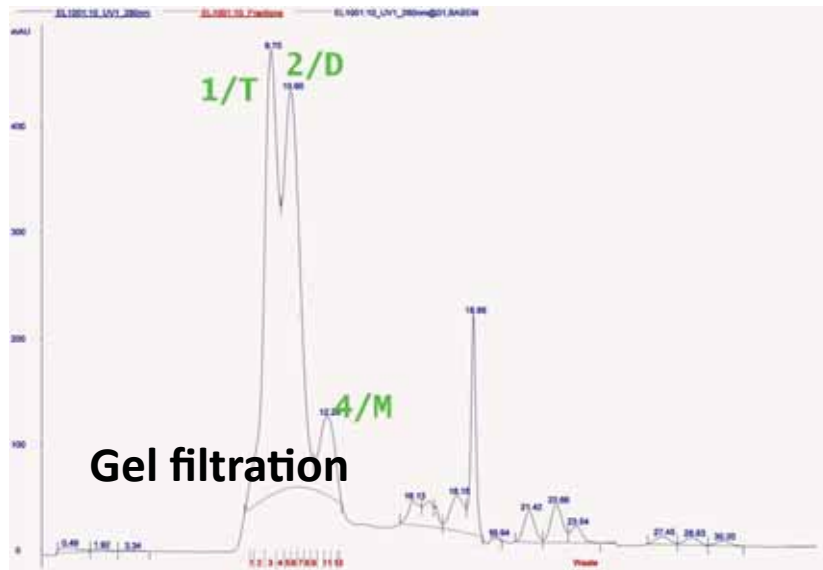
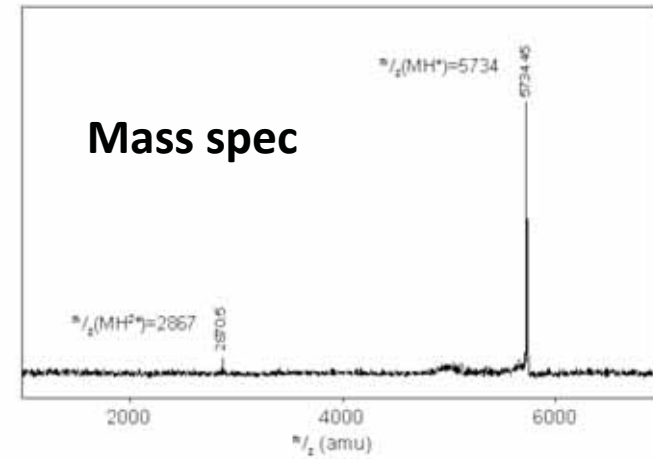
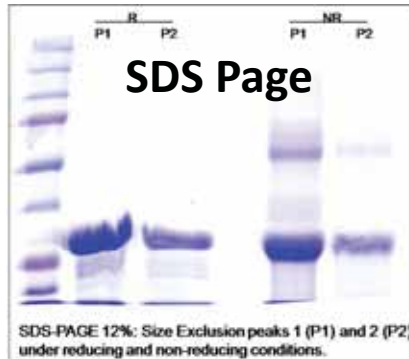
# Sample preparation and characterization around SAXS

Rob Meijers  
EMBL Hamburg

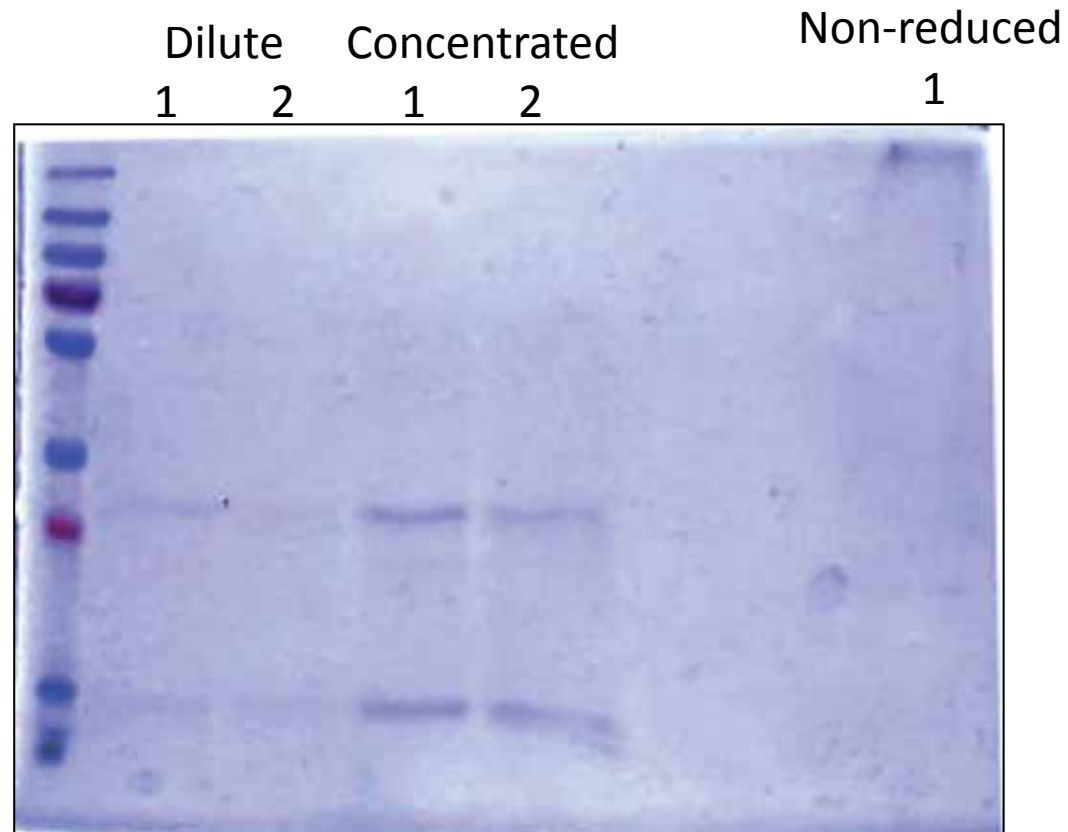
# Garbage in



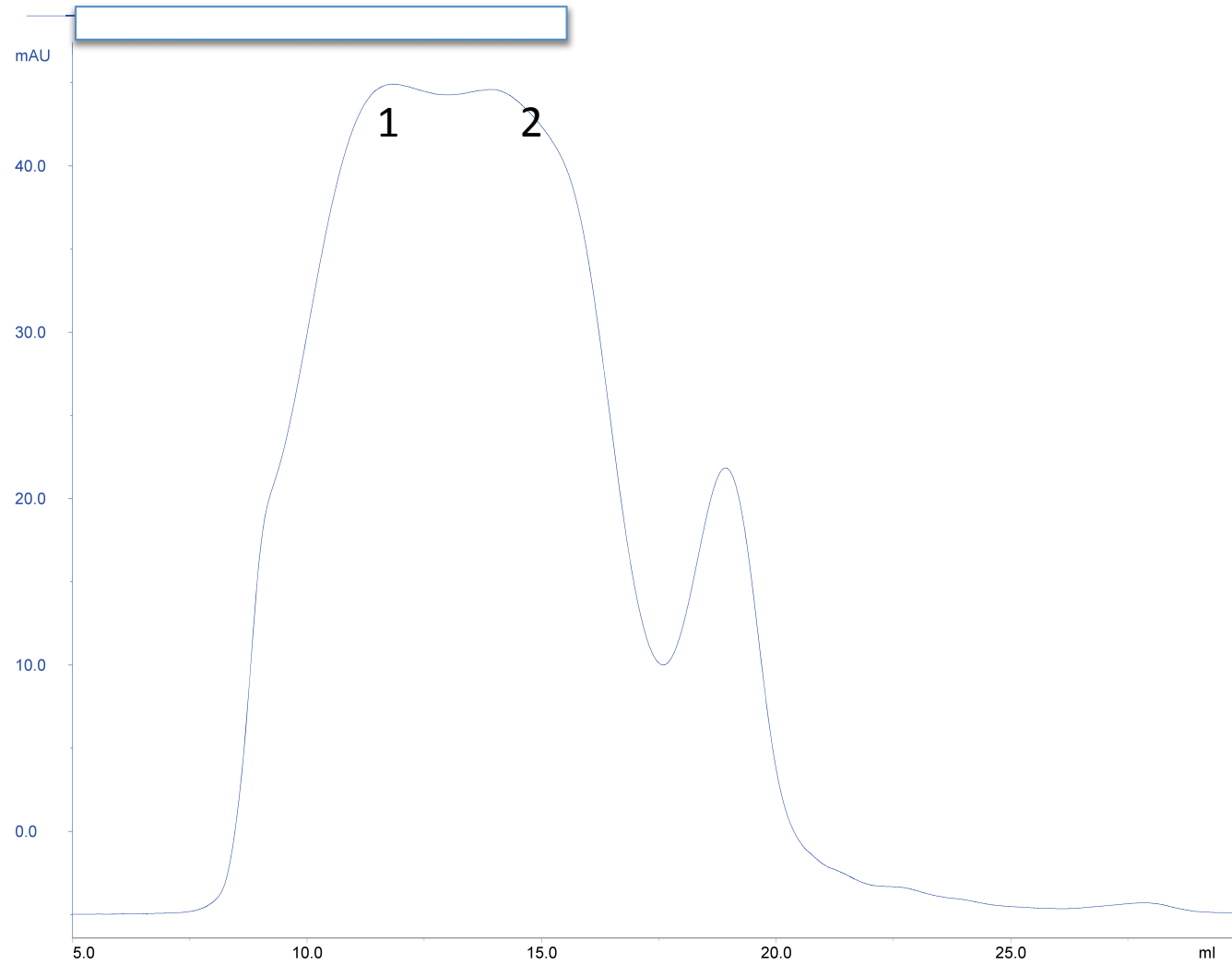
# Quality control



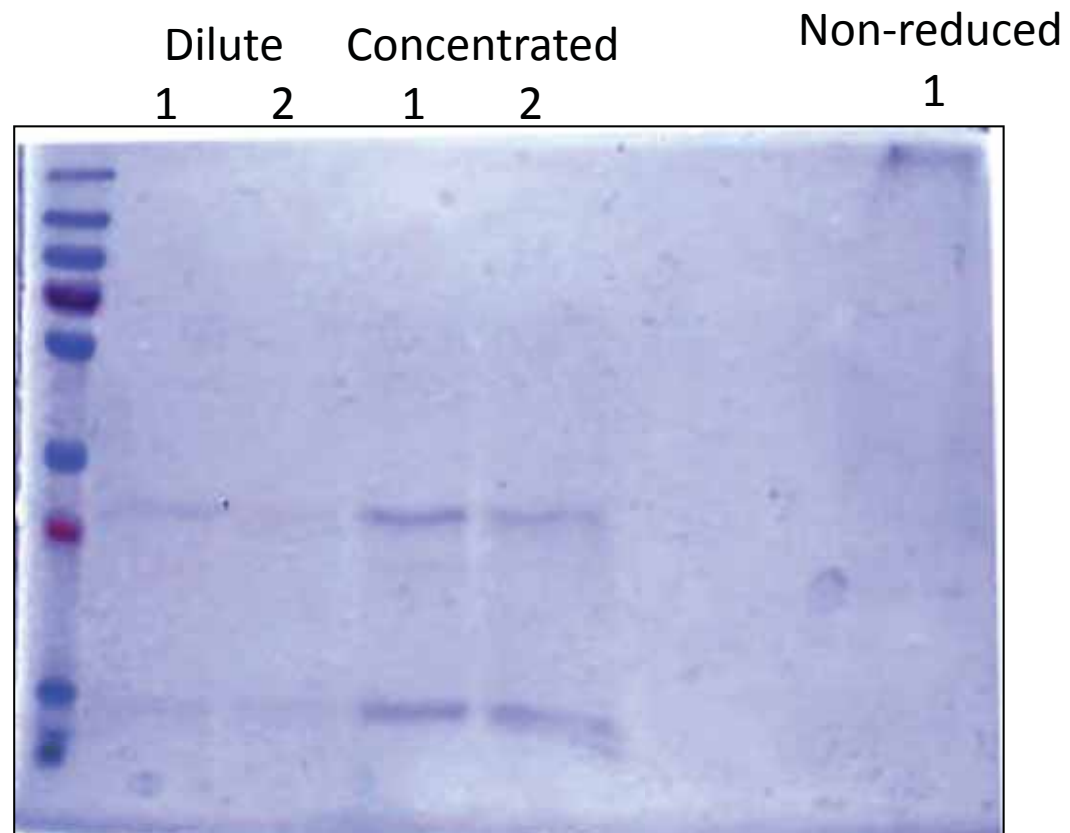
# A gel of a heterodimeric receptor



# A size exclusion profile



# A second look

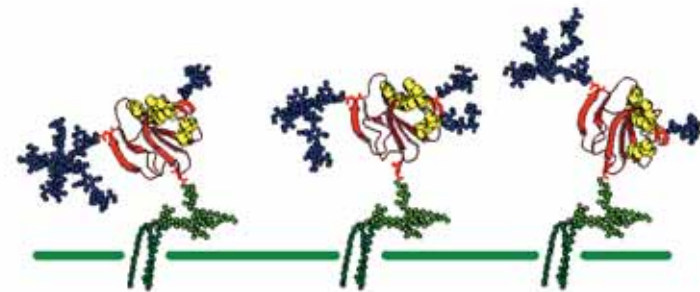


# Different gels

- SDS
- SDS non-reduced
- Native
- Iso-electric focussing
- Purity
- Disulphide bonds
- Post-translational modifications:
  - Phosphorylation
  - glycosylation

# Glycans form a sizeable part of the protein

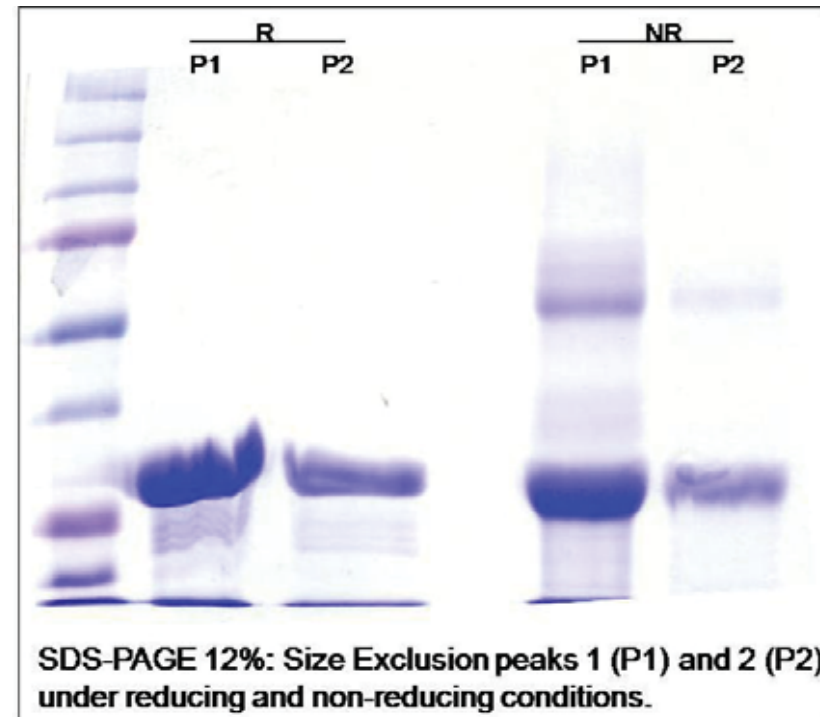
- Depends on expression system
- In pichia: > 40 % of MW can be glycans!
- Deglycosylate
- Change to HEK/COS/CHO-Lec



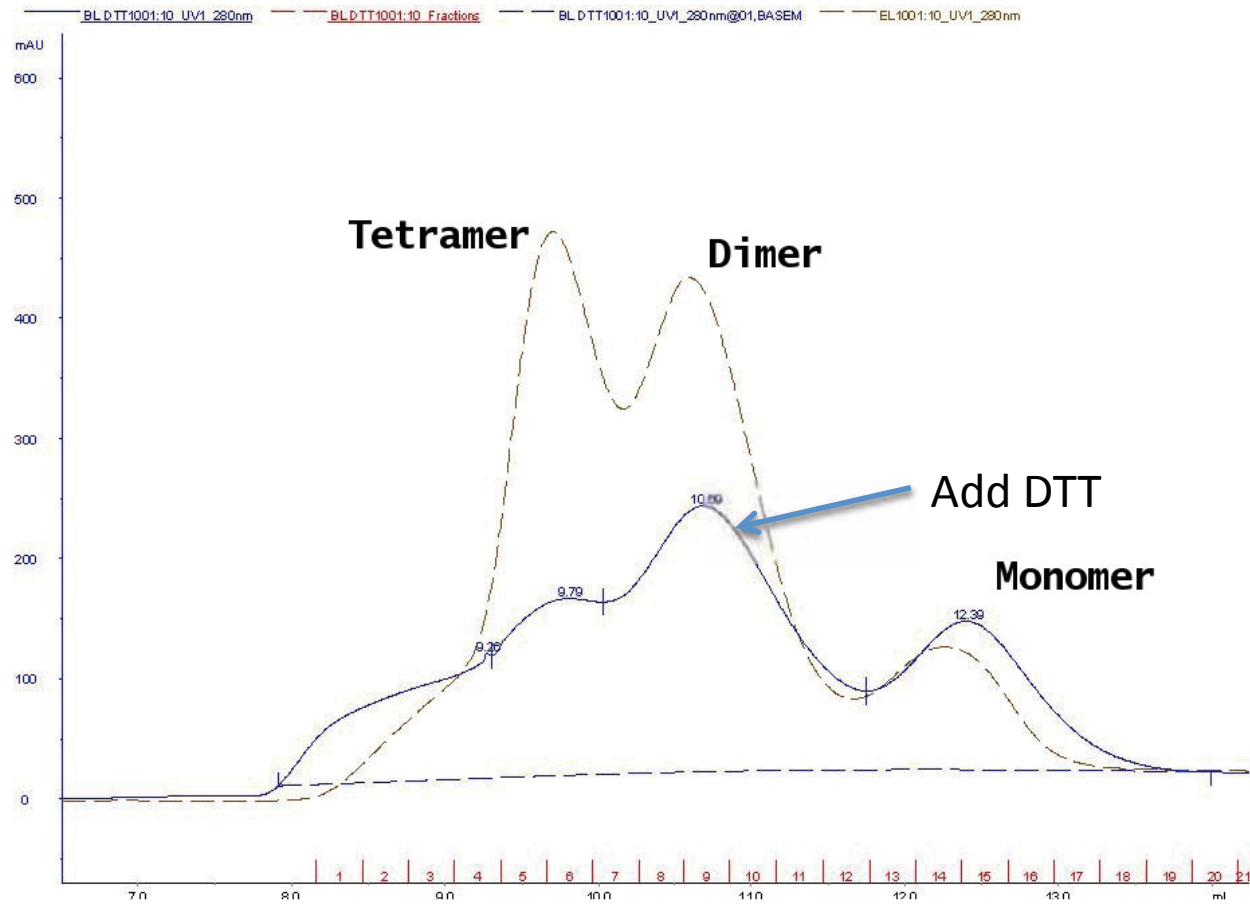
Oxford Glycobiology Institute

# Another capricious sample

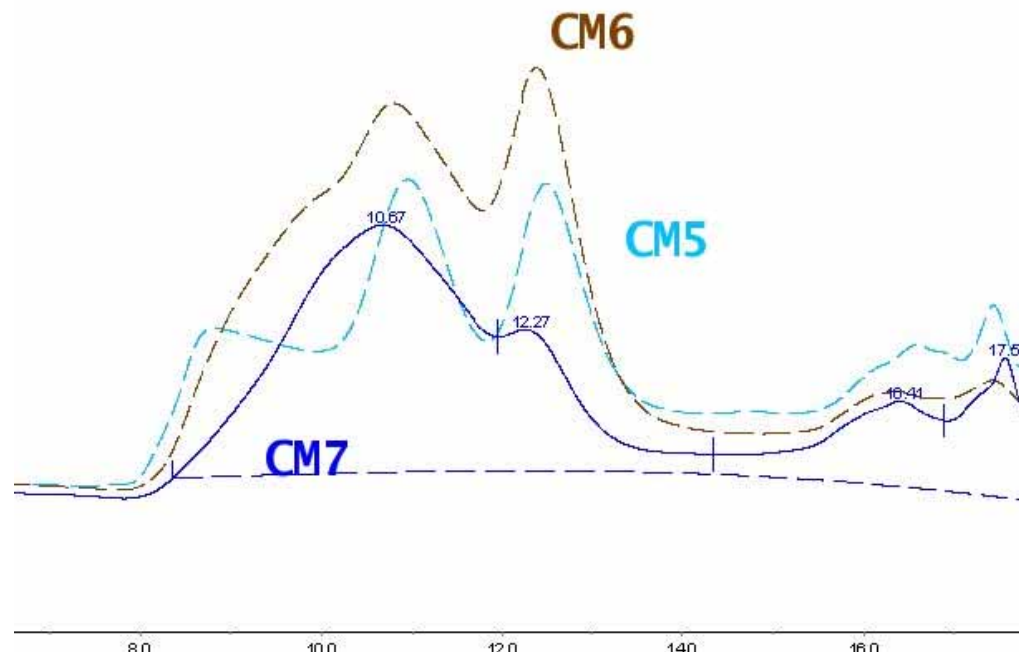
7XCYS ORIGINAL SEQUENCE, WITH HIS-TAG



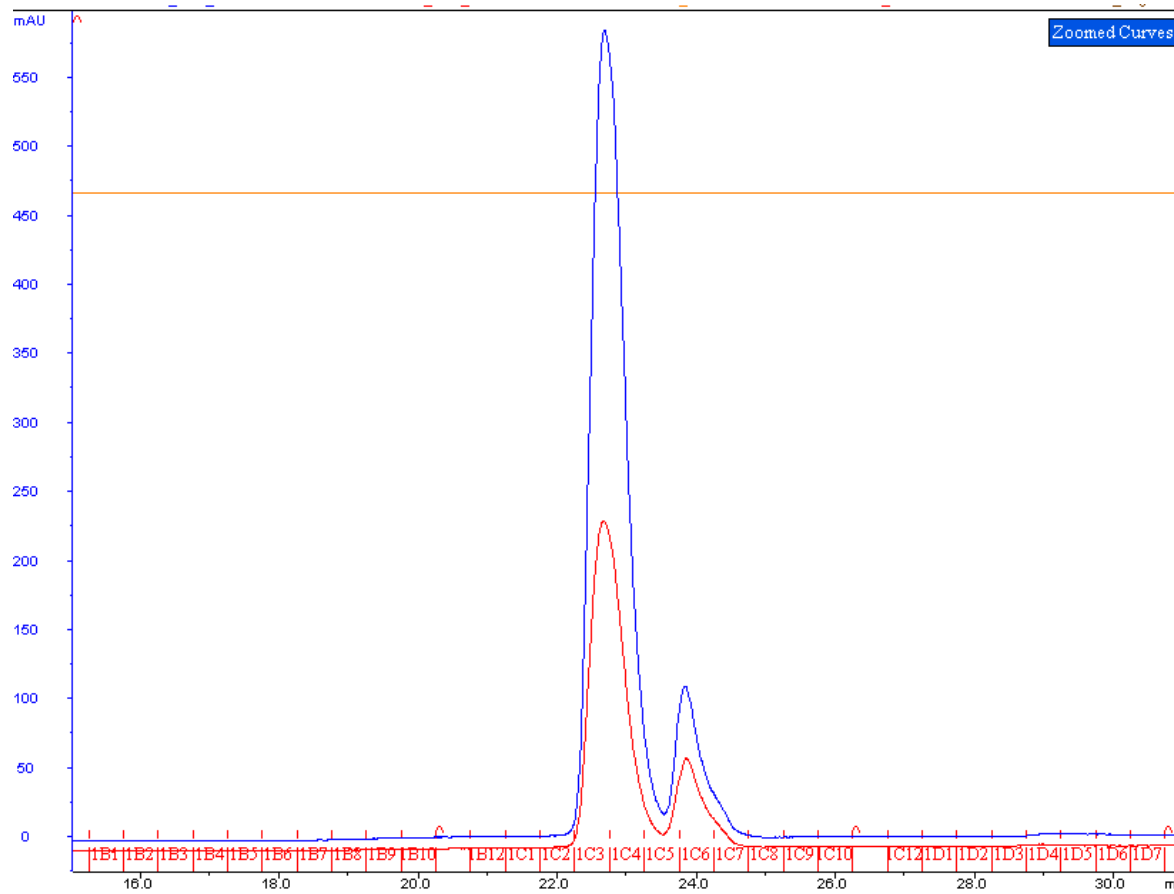
# Sizing profile



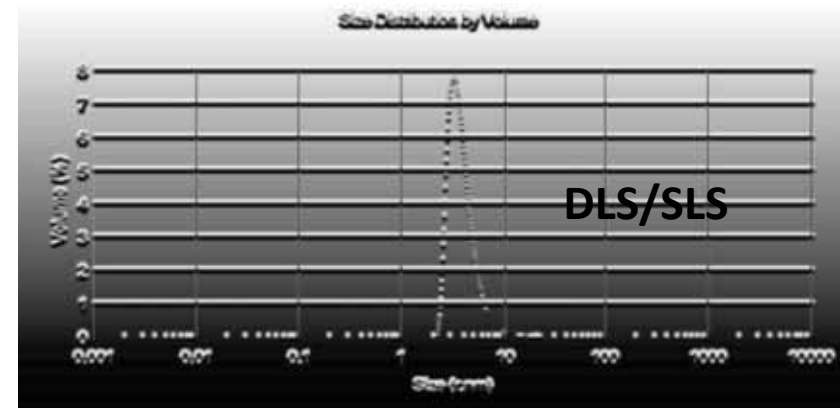
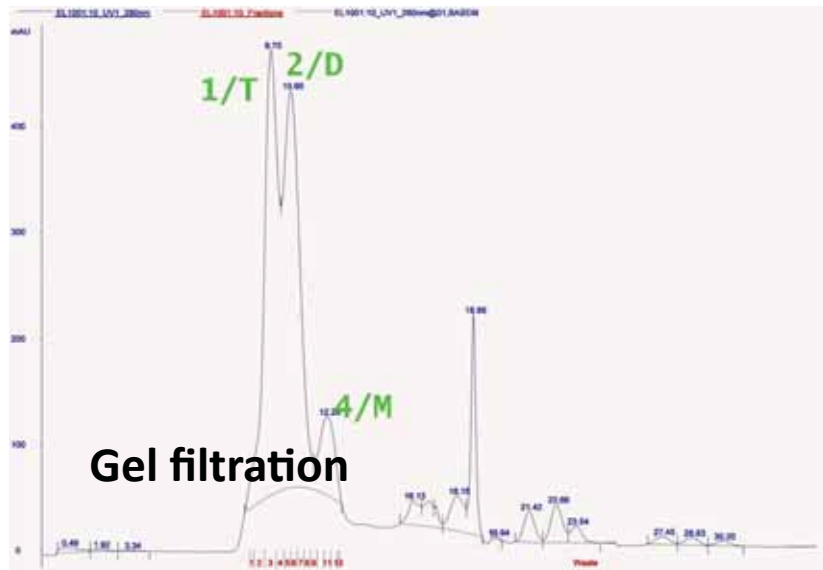
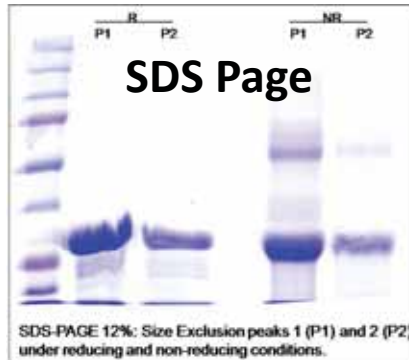
# Cysteine point mutations



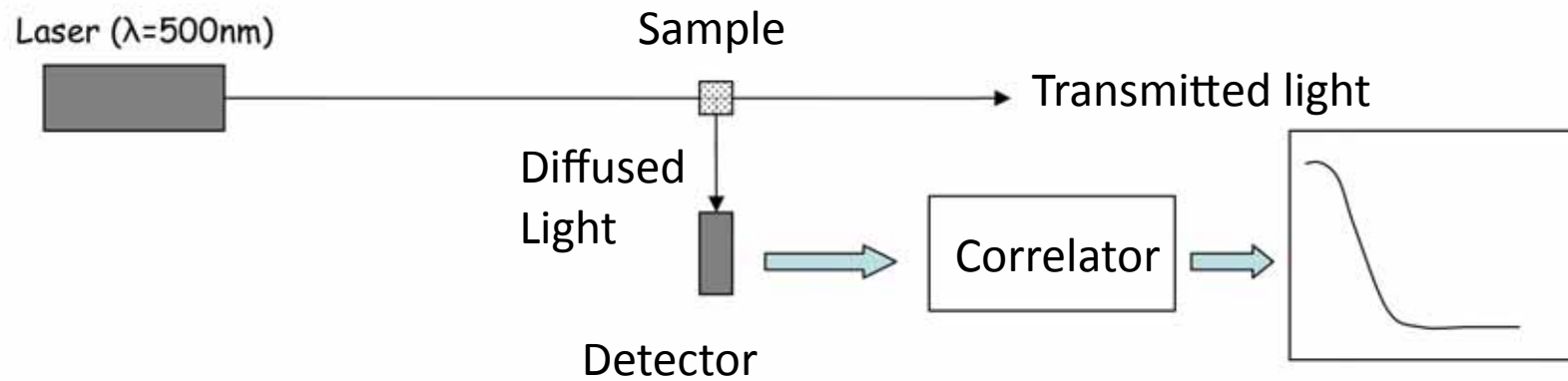
# Sizing profile



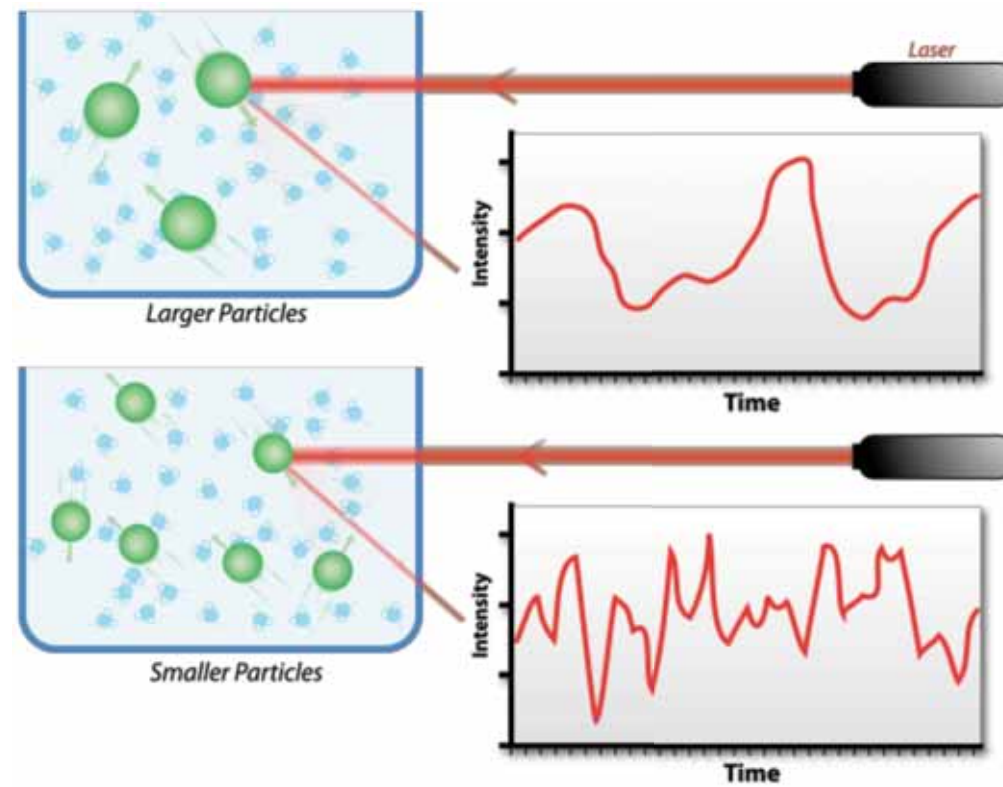
# Quality control



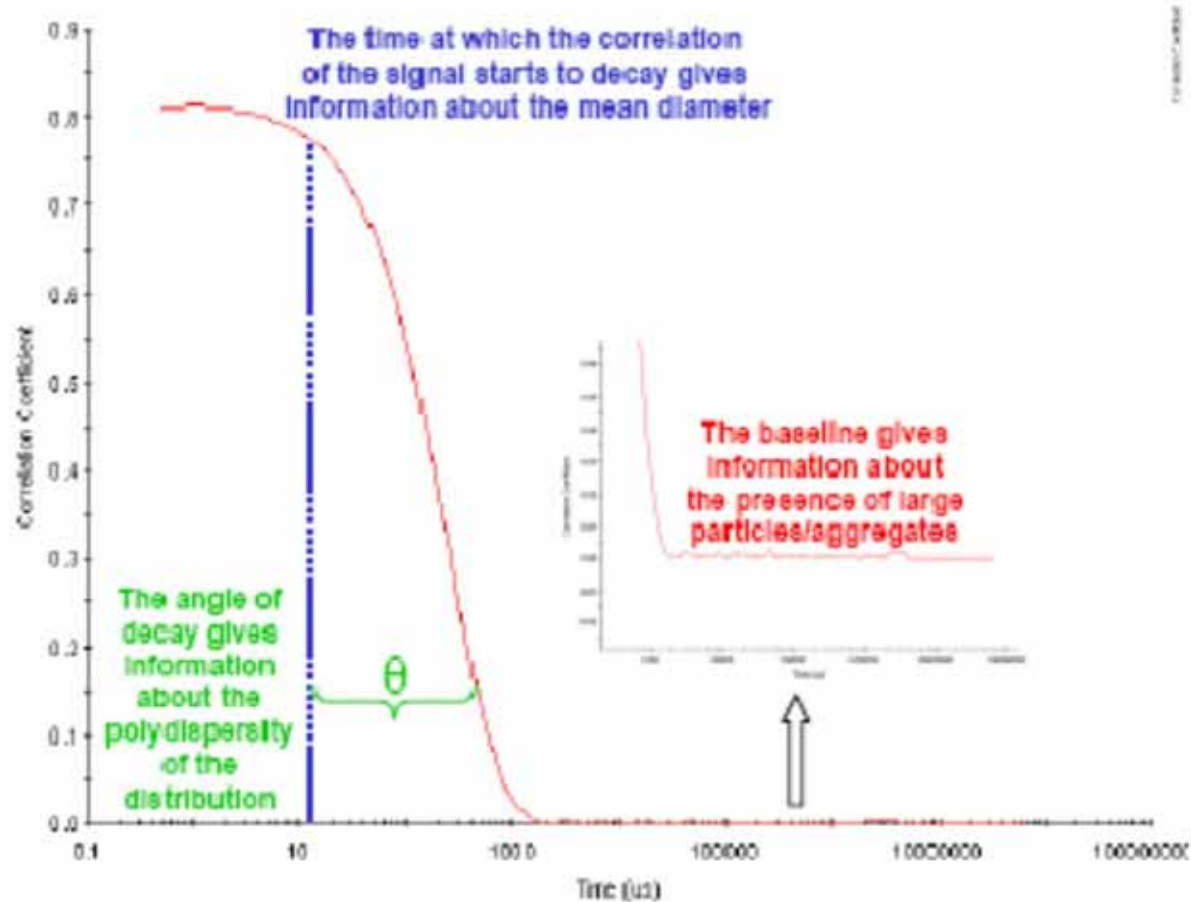
# Dynamic Light Scattering scheme



# Dynamic light scattering



# Information from the correlation curve



# Stokes-Einstein relation

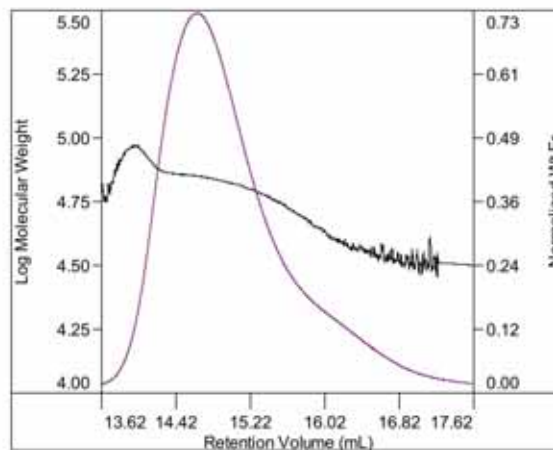
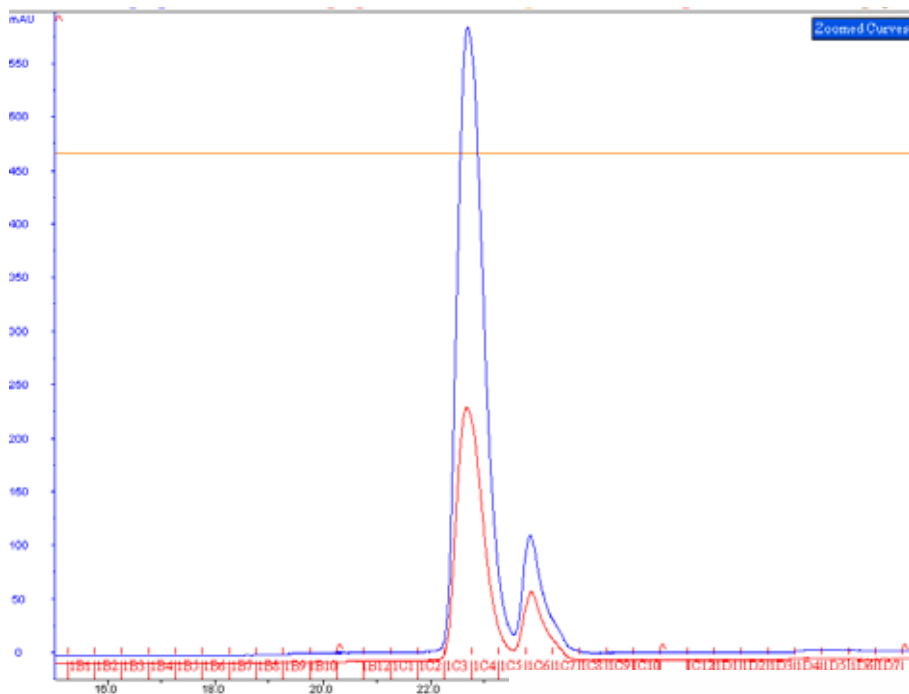
- $D$  = Diffusion coefficient
- $k$  = Boltzmann's coefficient
- $T$  = Temperature
- $\eta$  = Viscosity
- $R$  = hydrodynamic radius

$$D = \frac{kT}{6\pi\eta R}$$

# *Static Light Scattering*

Courtesy Bernd Tartsch, Malvern Instruments

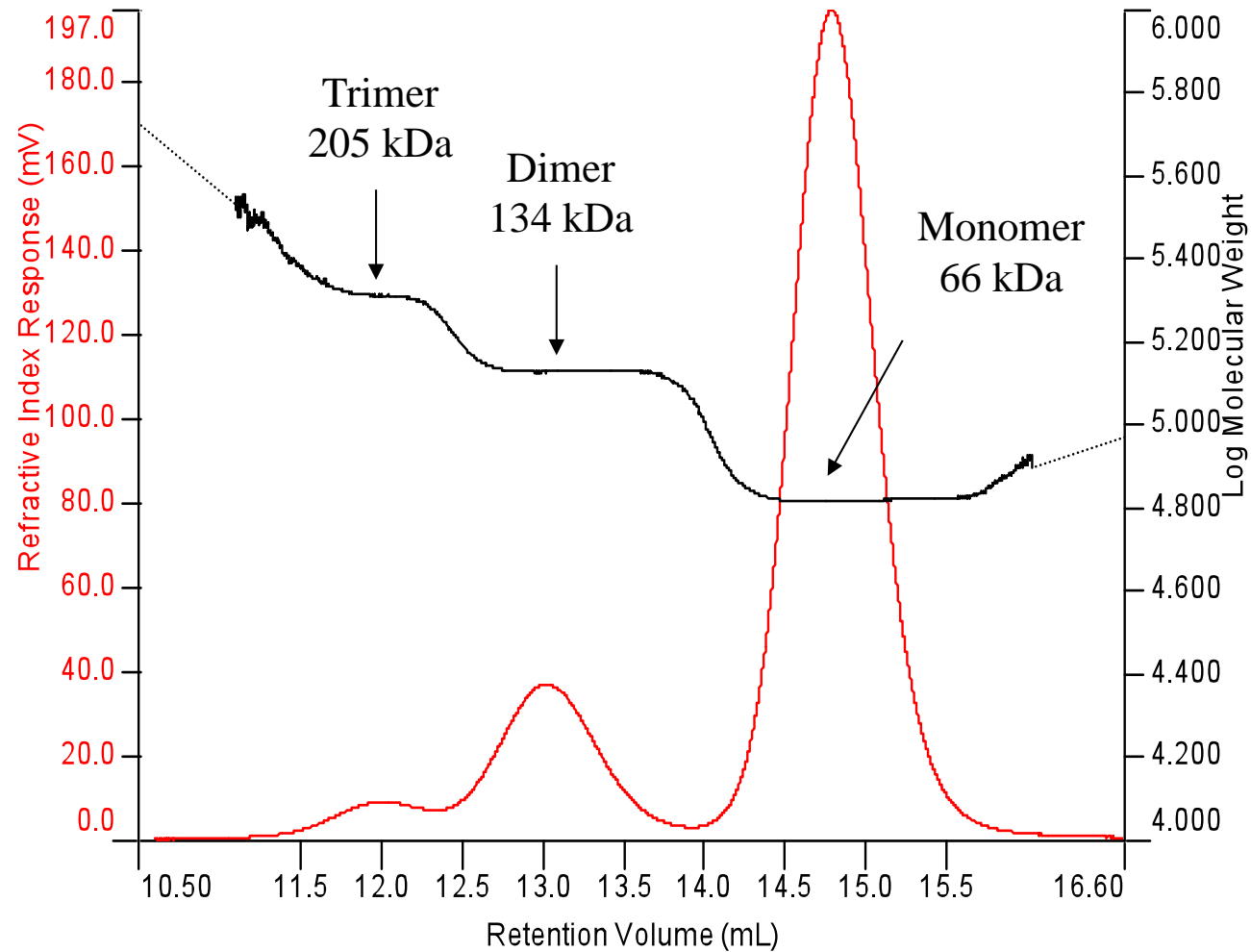
# Sizing profile, SLS twist



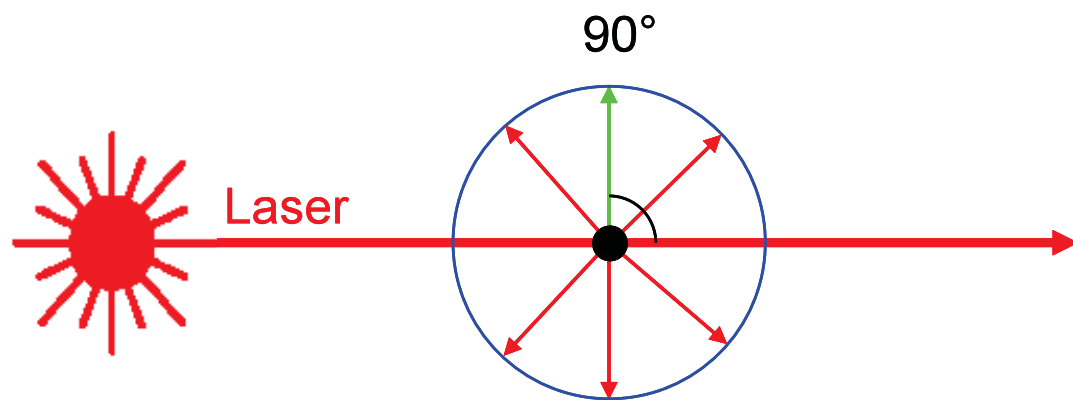
<b>Peak</b>	1		
<b>Mw (Daltons)</b>	64.151		
<b>Mw/Mn</b>	1.055		
<b>IV (dl/g)</b>	0.0434		
<b>Rh (nm)</b>	3.50		
<b>Wt Fr (Peak)</b>	1.0000		

<b>ID</b>	ctp11
<b>Solvent</b>	20mM HEPES, 150mM NaCl
<b>Flow Rate</b>	0.5000
<b>Conc (mg/ml)</b>	2.0079
<b>Inj Vol (ul)</b>	100.0

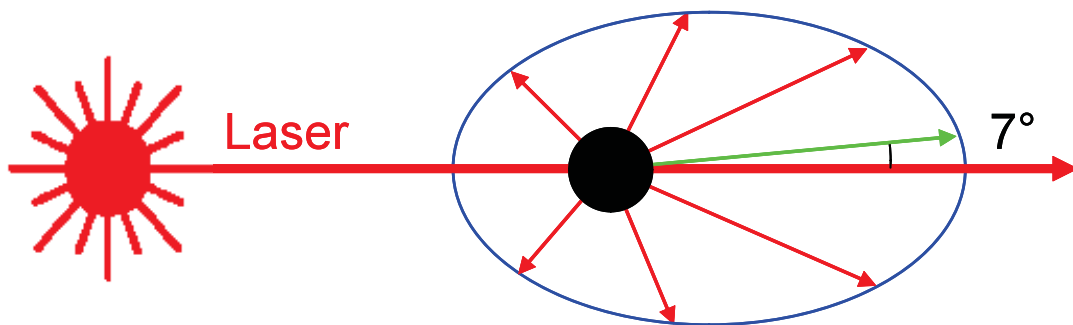
# BSA – Molecular Weight



# Angular Dependence

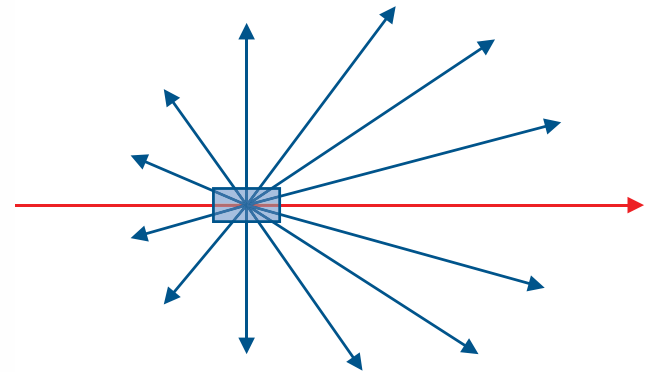
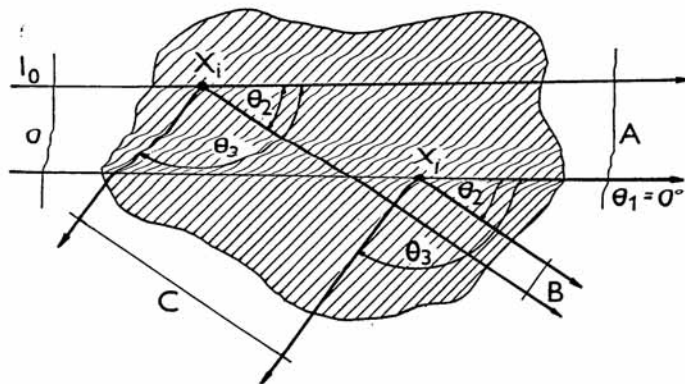


**small molecules**  
radius < 15 nm  
 $P_{\theta} = 1$  for all  $\theta$



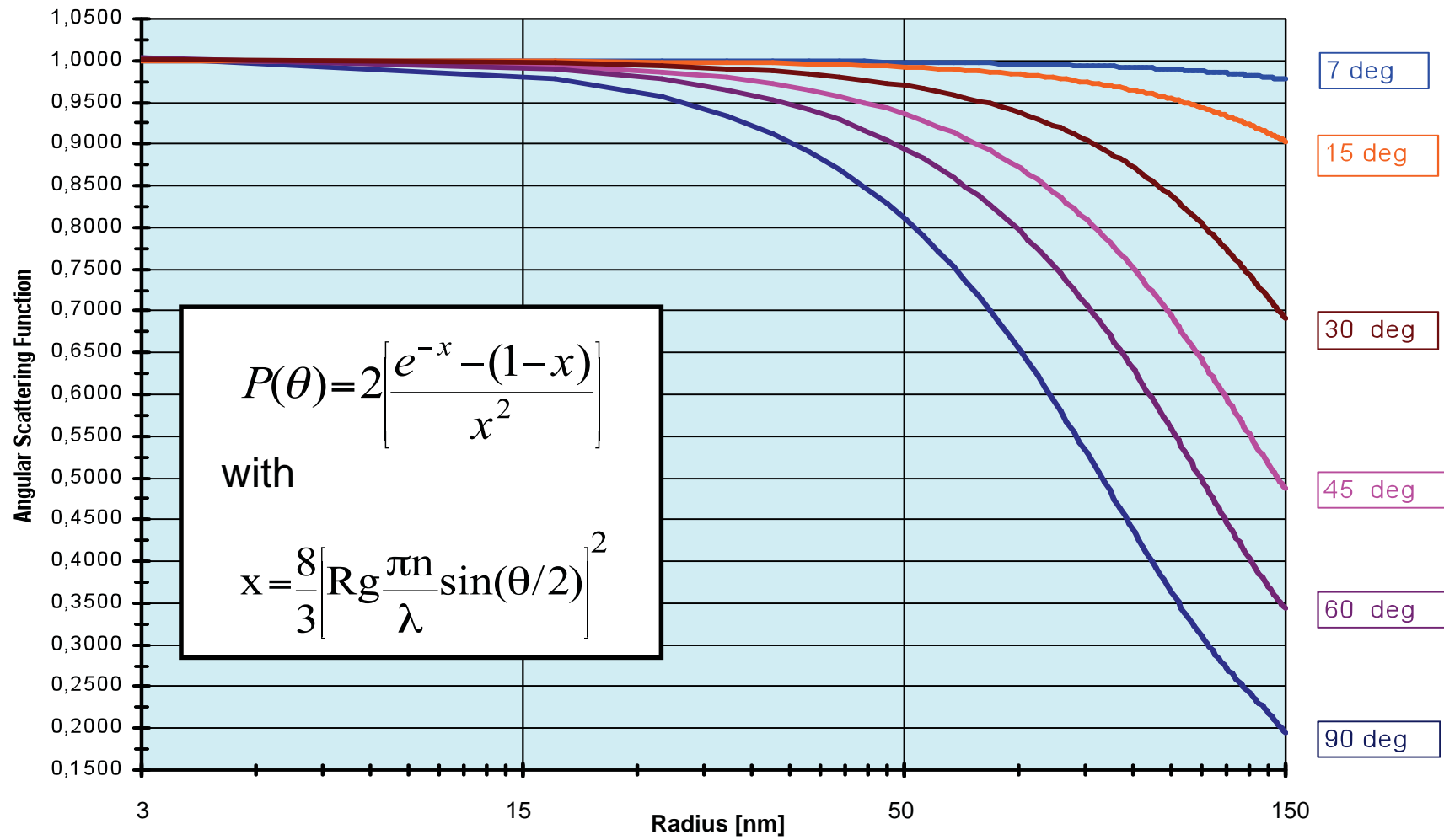
**large molecules**  
radius > 15 nm  
 $P_{\theta} = 1$  for  $\theta = 0^{\circ}$

Intramolecular interference produces a disymmetry in the scattered light.

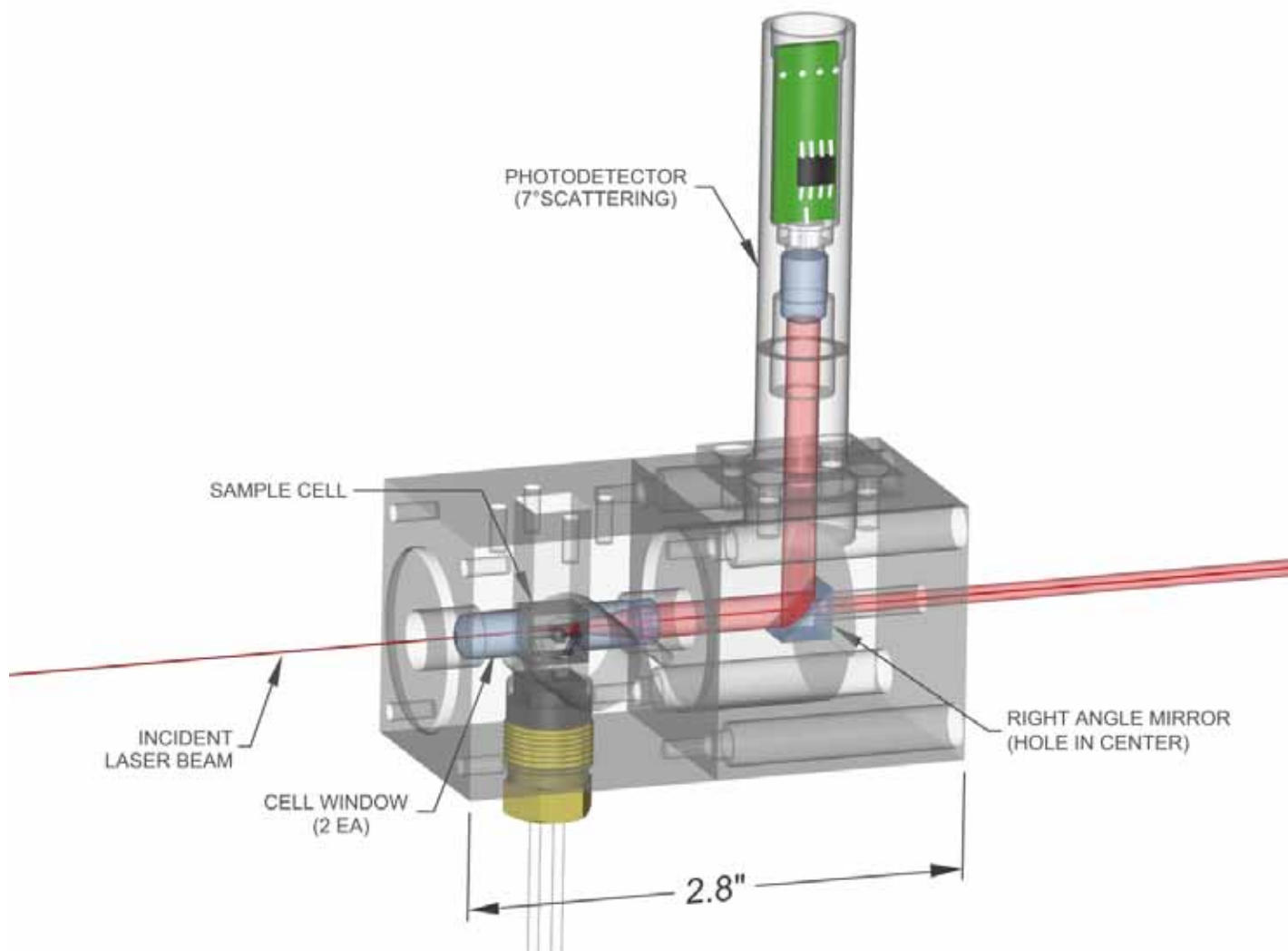


**Size of molecule/particle must be significant compared to wavelength of light**

# Angular Dependence



# Low Angle Light Scattering - LALS



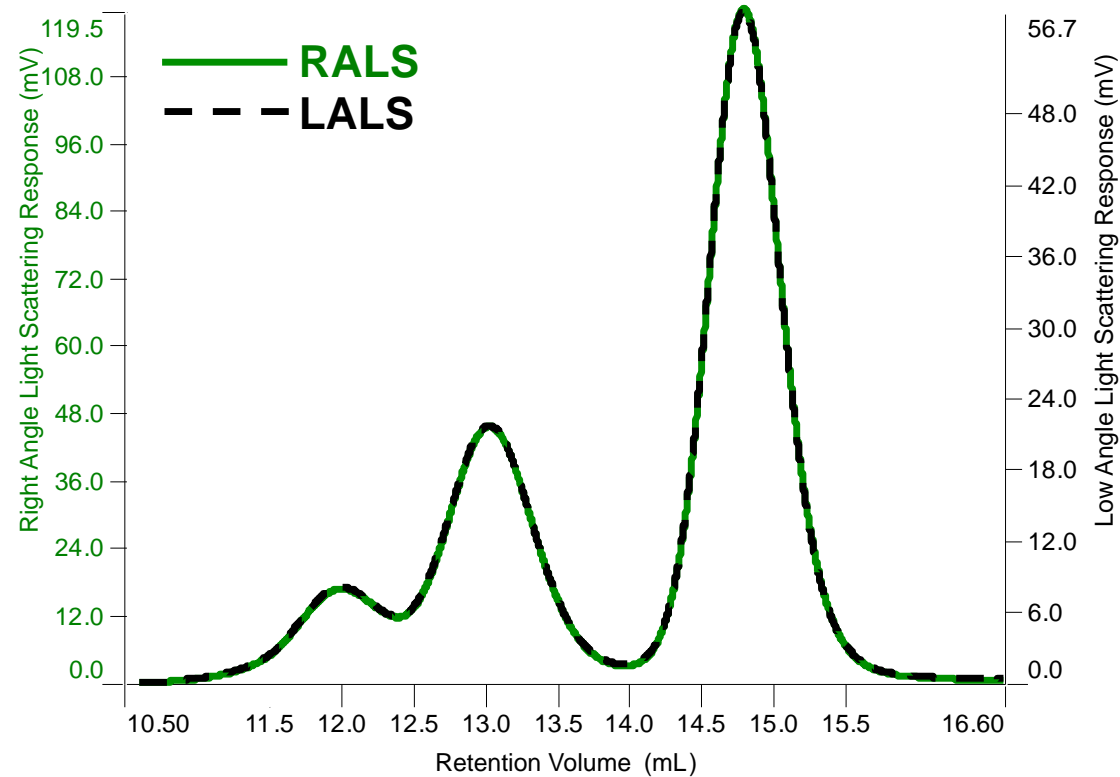
## Relationship between Size and Molecular Weight

<b>Proteins</b>		
	<b>Mol. Weight (D)</b>	<b>R<sub>h</sub> (nm)</b>
Ribonuclease	13,700	1.93
b-Lactoglobulin	35,000	2.67
Serum albumin	65,000	3.38
Hemoglobin	68,000	3.40
IgG	150,000	5.02
Catalase	250,000	5.38
Thyroglobulin	660,000	7.50
RALS		
-----		
<b>LALS</b>		
IgM	900,000	11.7
DNA	3 Mio	70

<b>Polysaccharides/PEG</b>	
<b>MW</b>	<b>R<sub>h</sub> (nm)</b>
12,000	2.62
50,000	5.62
80,000	7.27
150,000	9.81
RALS	
-----	
<b>LALS</b>	
270,000	12.74
410,000	15.07
670,000	18.45

# Normalised LS signals show no angular dependence for proteins

- Molecular weight requires only RALS
- Can not measure size by light scattering alone



*Addition of a  
Viscosity Detector  
gives  
hydrodynamic radius*

# Intrinsic Viscosity

Intrinsic viscosity is the concentration normalized viscosity of the sample in solution at infinite dilution.

$$IV = [\eta] = \left. \frac{\eta_{sp}}{c} \right|_{c \rightarrow 0} \quad \eta_{sp} = \frac{\eta - \eta_0}{\eta_0}$$

$\eta_{sp}$  is called the specific viscosity of the solution whose concentration is C.

$\eta_0$  is the Solvent Viscosity.

$\eta$  is the Solution Viscosity.

*In GPC, concentrations of solutions are sufficiently dilute that the extrapolation to zero concentration is negligible.*

# Hydrodynamic Radius

$$[\eta] = \frac{2.5}{\rho} = \frac{2.5 \cdot V_h}{M}$$

$$[\eta] M = 2.5 \cdot V_h = 2.5 \cdot \left( \frac{4}{3} \pi R_h^3 \right) = 10.47 R_h^3$$

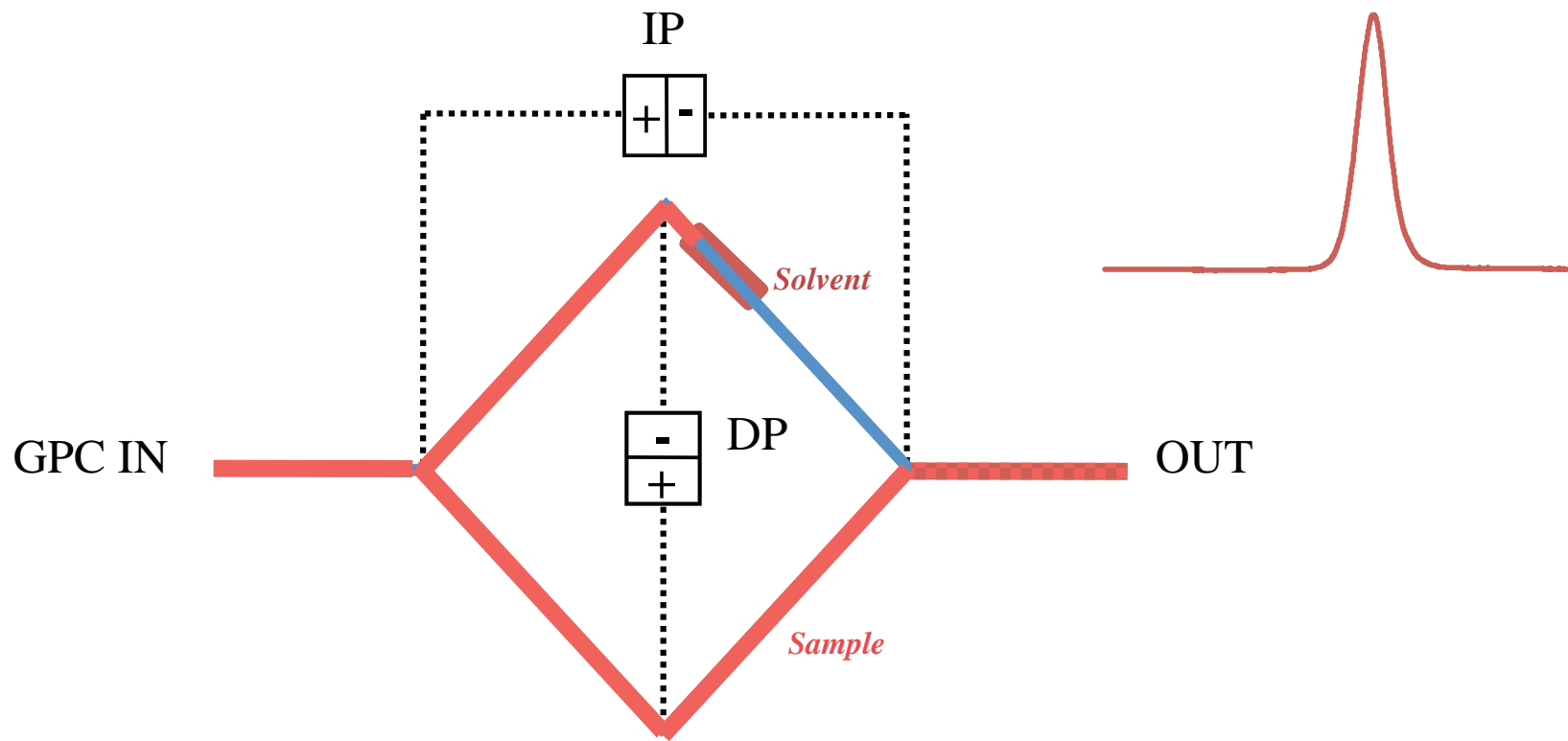
**Intrinsic Viscosity is inverse proportional to the density**

**Intrinsic Viscosity x Molecular weight is proportional to the hydrodynamic radius of a sample**

**Intrinsic Viscosity: from Viscosity Detector**

**Molecular Weight: from Light Scattering Detector**

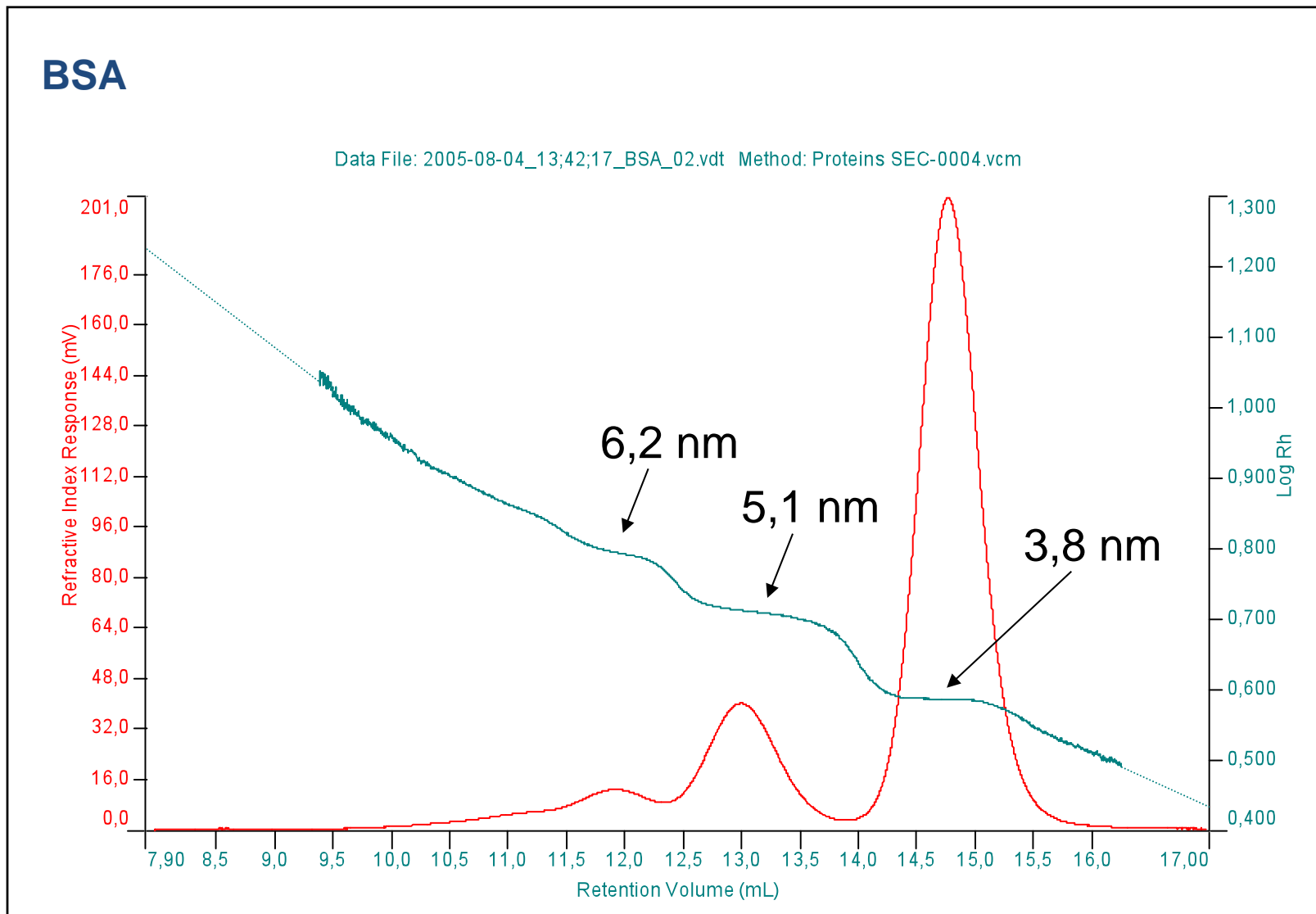
# Capillary Differential Viscosity Detector



Wheatstone bridge concept  
M. Haney, 1983

$$\eta_{sp} = \frac{4DP}{IP - 2DP} = C \times IV$$

# Hydrodynamic Radius



# Online Quality control

- UV
- Refractive index
- Viscosity
- Low angle light scattering



Viscotek/Malvern

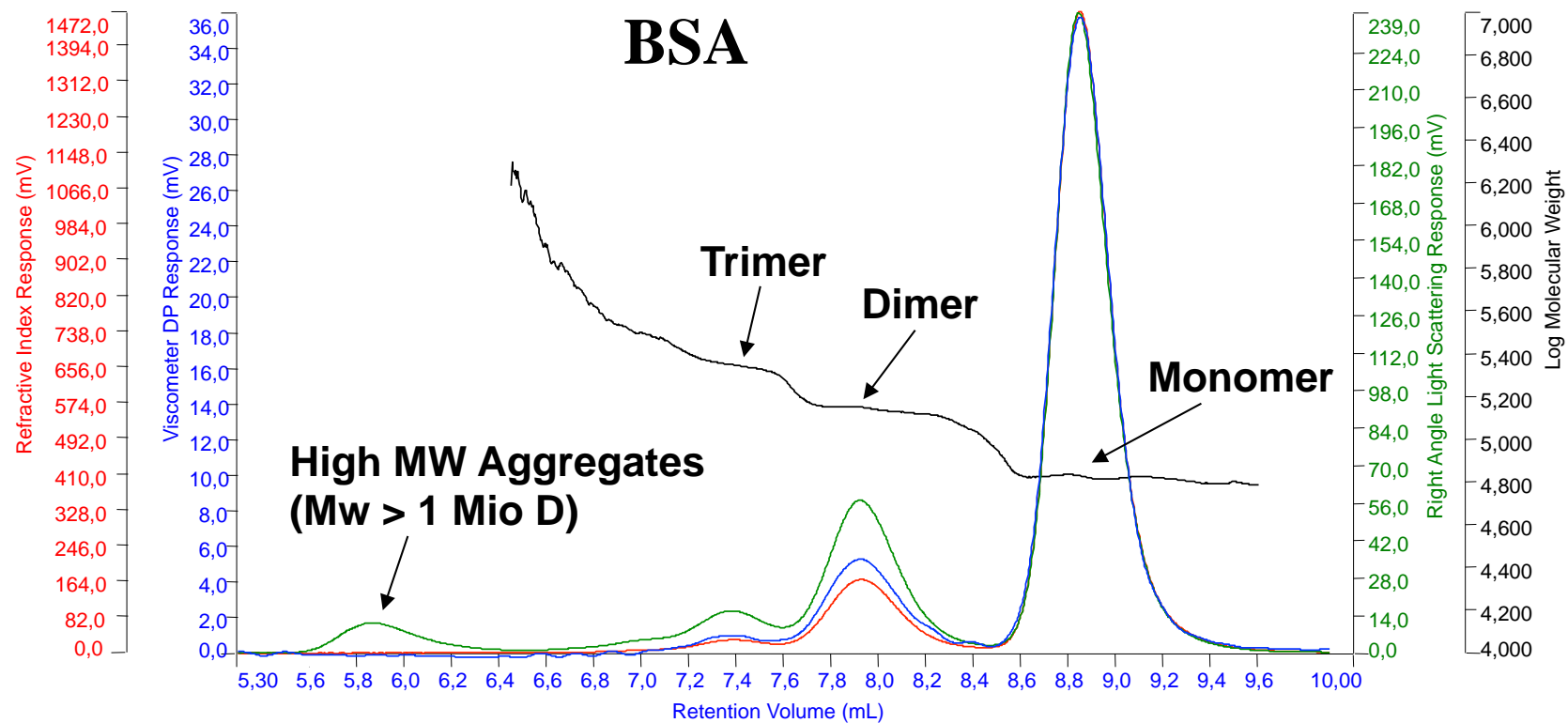
## What are the detectors responding to?

**Refractometer** =  $K_{RI} \times dn/dc \times \text{Conc}$

**UV-Detector** =  $K_{UV} \times dA/dc \times \text{Conc}$

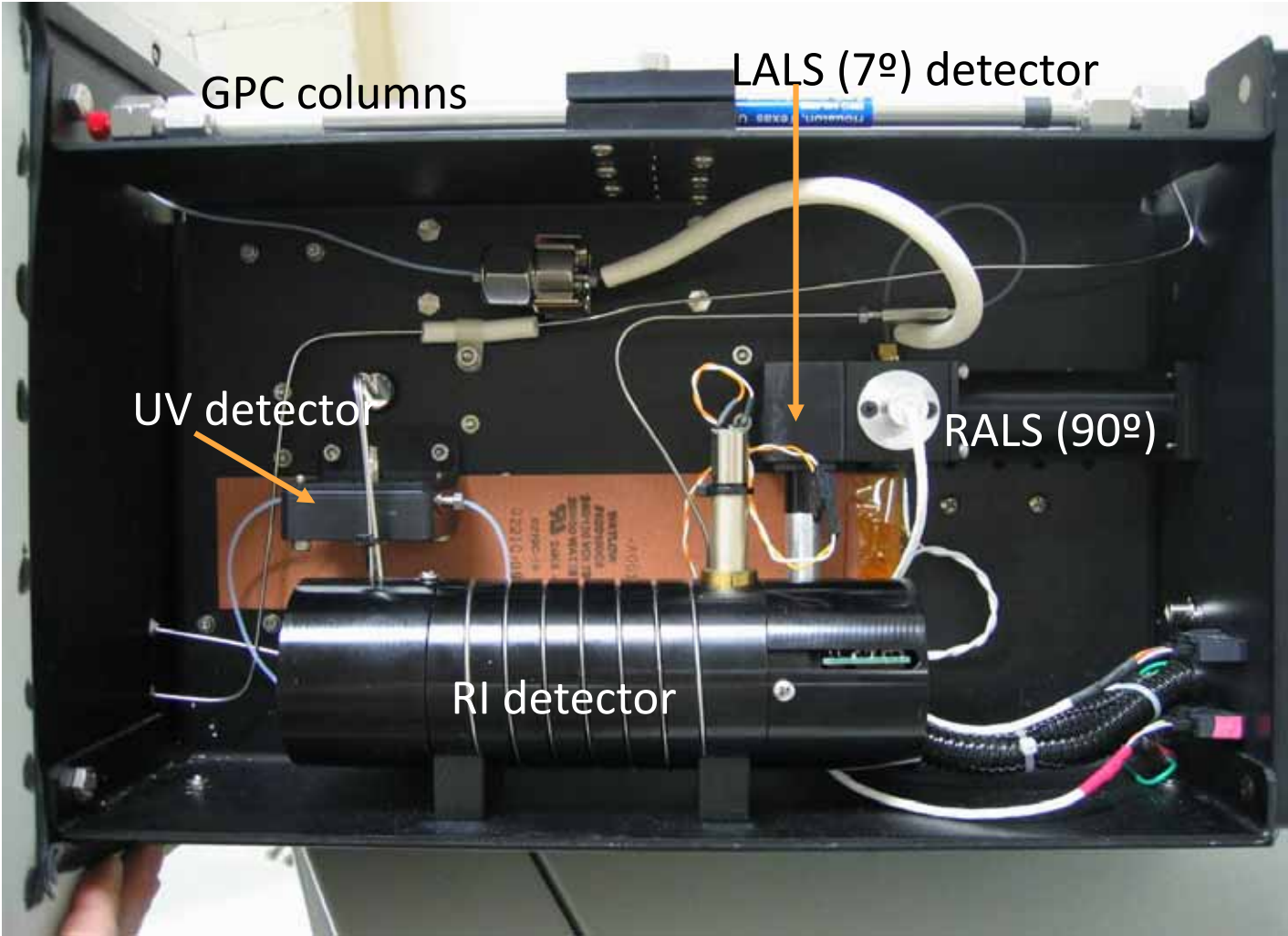
**Viscometer** =  $K_{Visc} \times \text{Intrinsic Viscosity} \times \text{Conc}$

**Light Scattering** =  $K_{LS} \times Mw \times (dn/dc)^2 \times \text{Conc}$



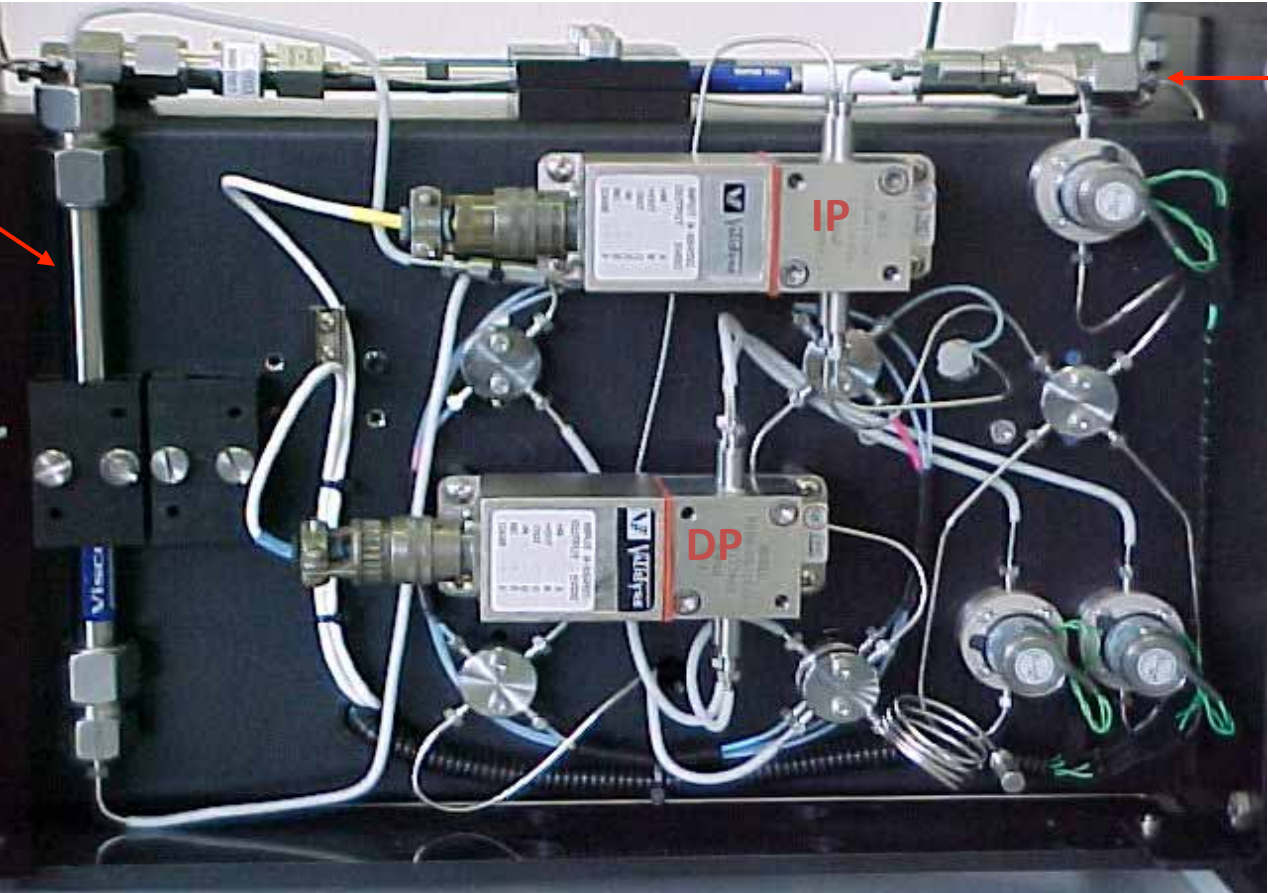
	Mw (D)	IVw (dl/g)	Rh (nm)	Weight Fraction (%)
<b>Monomer</b>	<b>66.430</b>	<b>0,056</b>	<b>3,88</b>	<b>87</b>
<b>Dimer</b>	<b>133.000</b>	<b>0,071</b>	<b>5,32</b>	<b>11</b>
<b>Trimer</b>	<b>201.000</b>	<b>0,095</b>	<b>6,69</b>	<b>1,5</b>
	<b>Light Scattering</b>	<b>Viscometer</b>		<b>Refractive Index</b>

# Laser Light Scattering Detector, Refractive Index Detector and UV-Cell



# Balanced Bridge 4-Capillary Viscometer Detector

Delay Volume



GPC Column(s)

# Online purification & QC



# Sample optimization

- Reduced Gel problematic: change purification protocol
- NR Gel problematic: check cysteines
- Protein aggregation, folding stability:
  - Size exclusion, light scattering, CD, NMR, thermofluor

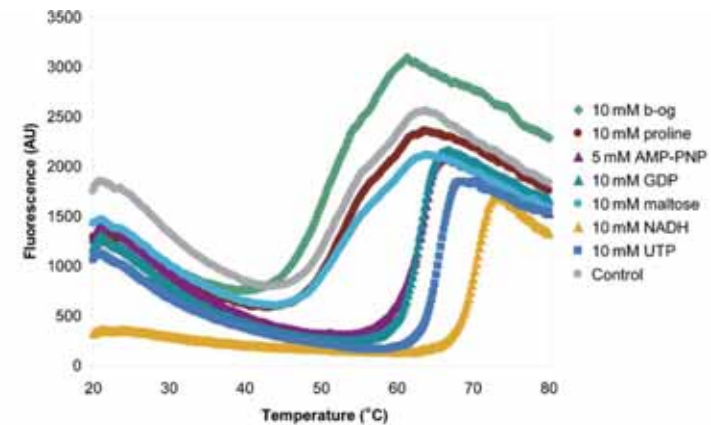
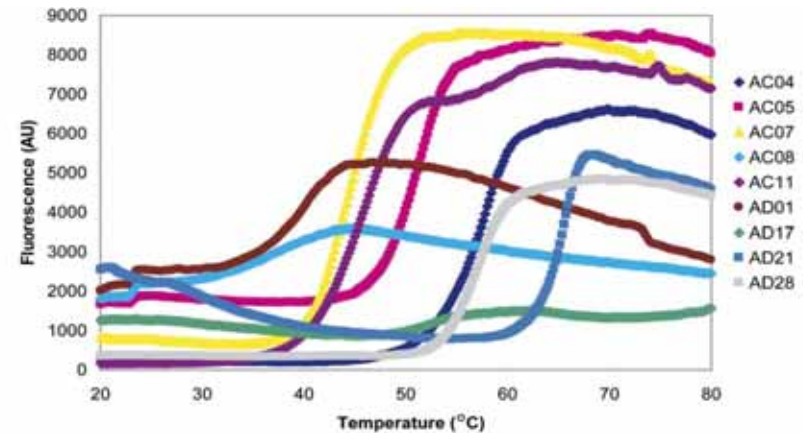
# Thermal stability

- ThermoFluor
- Modified real-time PCR machine
  - Add hydrophobic fluorescent probe
  - When protein unfolds...
  - Fluorescence increases



# Thermal stability

- Check protein stability
- Additive/ligand screen



# Sample optimization

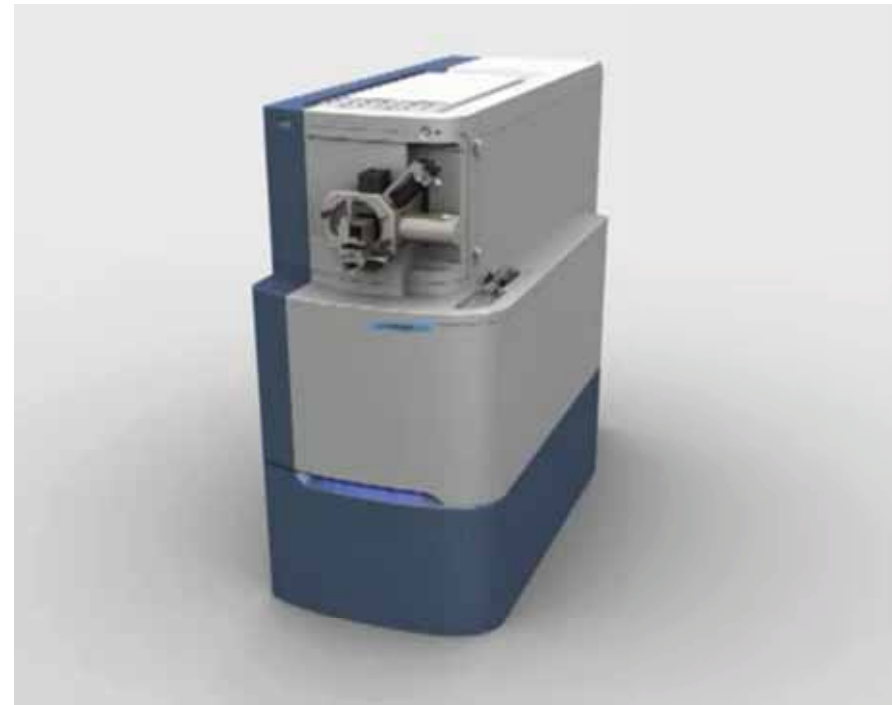
- Reduced Gel problematic: change purification protocol
- NR Gel problematic: check cysteines
- Protein aggregation, folding stability:
  - Size exclusion, light scattering, CD, NMR, **thermofluor**
- Modify buffers, additives
- If nothing works: change construct

# Additives?

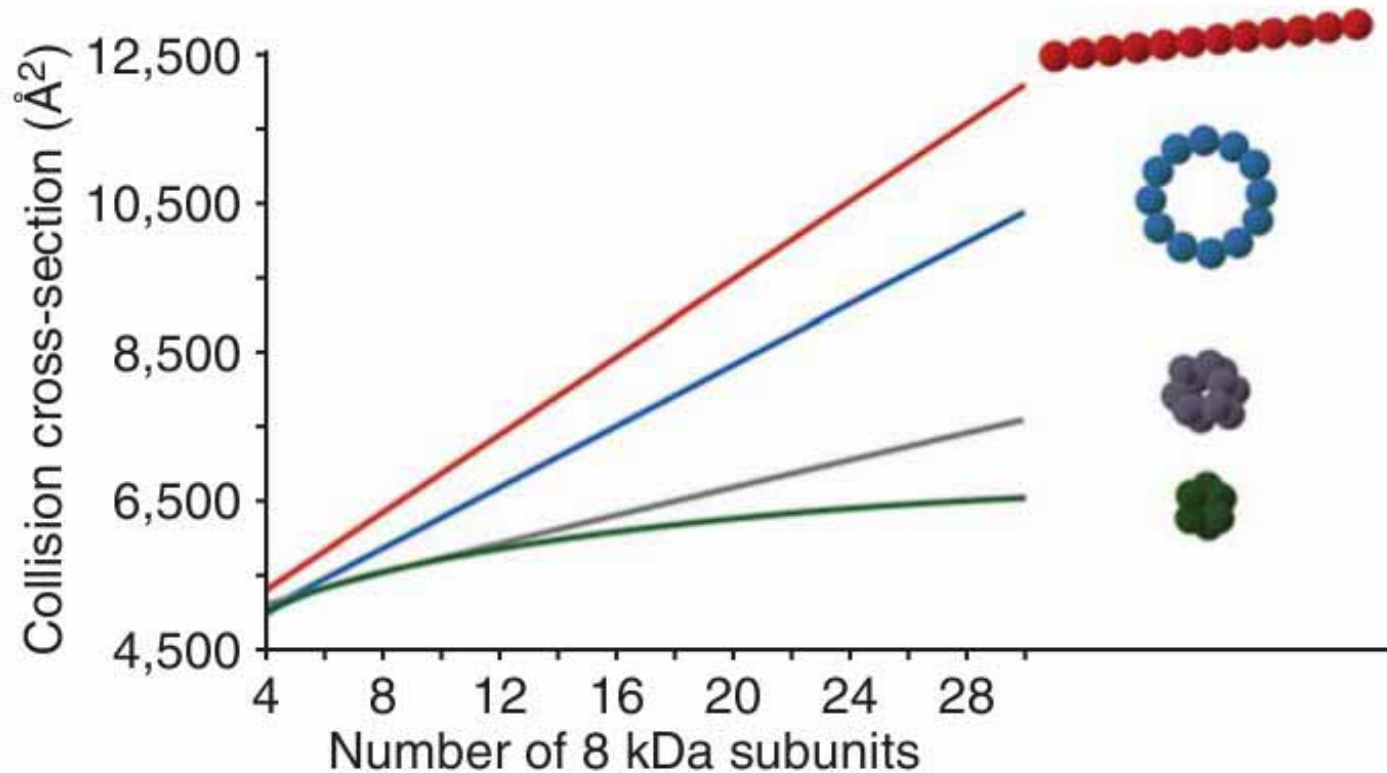
- **DTT**
- **Glycerol**
- **Detergents at less than 2xCMC**
  - 0.1% 1-s-Nonyl- $\beta$ -D-thioglucoside
  - 0.2% n-Decanoylsucrose
  - 0.3% n-Nonyl- $\beta$ -D-maltoside
  - 0.4% DDAO
  - 0.5% C8E5
  - 0.8% FOS-Choline<sup>®</sup>-10
  - 1.1% FOS-Choline<sup>®</sup>-9

# Mass spectrometry

- Traditional MS:
  - Confirm mass
  - Confirm sequence
  - Check folding state
- MS/MS + Ion mobility :
  - Detailed folding state
  - Protein-protein interactions
  - Whole protein size...



# Ion mobility derived particle size



Ruotolo et al *Nature Protocols* (2008) **3**, 1139

in combination with SAXS...

# Conclusions

- QC at home is crucial
- But soon we hope to do it for you at EMBL@PETRA3
- Some quality control methods can provide useful complementary data
- This all can filter the garbage “in” bit

But the garbage out?



# Acknowledgements

- Malvern Instruments  
Bernd Tartsch